Amendments to the Specification:

Please remove the Sequence Listing filed March 26, 2001 and insert therefor the enclosed substitute Sequence Listing.

Please amend the paragraph spanning pages 9 and 10 as follows:

(Example 1: Activation of Tos17 via culture)

Using fully ripened seeds of Nipponbare and Akitakomachi, which are varieties of Japonica subspecies, induction of calli and cell suspension culture were carried out as described earlier (Hirochika et al., 1996, supra). The activation of Tos17 which was used for gene destruction was carried out following the method of Ohtsuki (1990) (~~rice protoplast culture system,~~ Yoshiaki Ohtsuki, "Video Manual for Experiments in Rice Protoplast Culture System," published by Food and Agricultural Research Development Association, Tokyo, Japan (1990)). In summary, fully ripened seeds of rice were cultured in an MS medium having 2,4-dichlorophenoxyacetic acid (2,4-D) added thereto (2 mg/ml) (Ohtsuki (1990), supra) (25°C, 1 month), to induce callus formation. The resultant calluses were cultured for 5 months in an N6 liquid medium having 2,4-D added thereto (Ohtsuki (1990), supra), and thereafter placed on a redifferentiation medium (Ohtsuki (1990), supra), whereby redifferentiated rice plants were obtained (first generation (R1) plants).

Please amend the paragraph spanning pages 10 and 11 as follows:

(Example 2: Isolation of sequences adjoining Tos17)

Utilizing each of the regenerated R1 rice plants obtained according to Example 1 as a first strain, about 1000 R1 seeds were collected from each strain and grown on a paddy field to obtain second generation (R2) plants, which were subjected to a morphological analysis. As a result of observing the phenotypes of the respective plant bodies in the R2 group, it was learned that about 1/4 of the R2 group of an Akitakomachi strain A0369 exhibit the "dwarfism, upright form, and malformation of grain hulls" phenotype (Figures **1A** and **1B**). In the regenerated group of Akitakomachi, dwarfism, upright form, and malformation of grain hulls were observed for brassinosteroid insensitive mutants (Figure **1A**, left, and Figure **1B**, left), as compared with the wild type (Figure **1A**, right, and Figure **1B**, right). The isolation of adjacent sequences of transposed Tos17, which is ~~co-segragating~~ co-segregating with the phenotypes, was carried out by an IPCR method (Ochman et al., Genetics Nov; 120(3): 621-3(1988) and Triglia et al., Nucleic Acids Res Aug 25; (16): 8186(1988)). The total DNA of A0369 was digested with XbaI, and a ligation process was performed in a large quantity of solution, thereby obtaining self-ligated circular molecules. In the self-ligated circular molecules, the adjacent sequences are flanking the internal sequence of Tos17. As a result, amplification was successfully carried out by usual PCR methods using an outward primer pair (T17TAIL3: GAGAGCATCATCGGTTACATCTTCTC (SEQ ID NO: 4); T17-1950R: TCTAGCAGTCTCAATGATGTGGCG (SEQ ID NO: 5)) based on the known sequence of Tos17.